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## Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

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**To cite this Article** Lewis, Maureen , Brian, T. , McMurry, H. and De Clercq, Erik(1995) 'Fluorinated Carbaacyclonucleosides: Synthesis and Evaluation of Antiviral Activity', *Nucleosides, Nucleotides and Nucleic Acids*, 14: 9, 1913 – 1927

**To link to this Article:** DOI: 10.1080/15257779508010714

**URL:** <http://dx.doi.org/10.1080/15257779508010714>

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**FLUORINATED CARBAACYCLONUCLEOSIDES:  
SYNTHESIS AND EVALUATION OF ANTIVIRAL ACTIVITY.**

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*Abstract:* Two series of fluoroacyclic nucleosides were synthesised by condensation of nucleic acid bases with substituted fluoropentanes. 1-(5'-Fluoro-4'-hydroxypentyl)-cytosine showed a modest activity against cytomegalovirus (MIC<sub>50</sub>: 12-15 µg/ml). All the other compounds were inactive against all the viruses tested.

The discovery of the antiviral potency of 9-[(2-hydroxyethoxy)methyl]guanine (acyclovir)<sup>1</sup> stimulated the synthesis of a large number of acyclonucleosides with various side chains and aglycones<sup>2,3</sup>. Carbocyclic analogues of nucleosides in which the oxygen of furanose is replaced by a methylene group show increased resistance to phosphorylases while maintaining their biological activity.<sup>4</sup> Replacement of a hydrogen atom by fluorine does not produce drastic steric changes in a molecule<sup>5a</sup> but may enhance the biological activity of the molecule. The much greater electronegativity of fluorine has a strong electronic effect on reactions of neighbouring functional groups and the fluorine atom, being a hydrogen bond acceptor, is often processed by enzymes acting on the corresponding hydroxy compounds.<sup>5b</sup> Pyrimidine and purine nucleosides containing fluorine in the C2' (β) (arabino) configuration show dramatically different biological activity and enhanced chemical and metabolic stability compared to the parent deoxynucleoside.<sup>6</sup> As an extension of previous work<sup>7</sup> we describe the synthesis of two series of fluorinated carbaacyclonucleosides.

### Chemistry

The alkylating moieties, 1-fluoro-5-iodopentan-2-yl acetate **1** and 5-fluoro-4-tosyloxypentyl pivalate **2** were obtained through ring opening of 2-(fluoromethyl) tetrahydrofuran as described previously<sup>7</sup> for the preparation of 2-amino-6-chloro-9-(1'-fluoro-5'-pivaloyloxypentan-2'-yl)-9H-purine **6** and 9-(4'-acetoxy-5'-fluoro-pentyl)-2-amino-6-chloro-9H-purine **12**.

The reaction of **2** with 4-methoxy-5-methyl-2-pyrimidinone in dimethylformamide using potassium carbonate as base and 18-crown-6 as catalyst<sup>8</sup>, gave a mixture of isomers (see below) from which the required product, **3** was isolated by chromatography. The cytosine analogue **4** was also obtained in the same way, but the yield was improved by use of cesium carbonate in place of potassium carbonate and 18-crown-6<sup>9</sup>. 1-(1'-Fluoro-5' -pivaloxypentan-2'-yl)-O<sup>4</sup>-methylthymine **3** was obtained in 30% yield and the isomeric O<sup>2</sup>-alkylated product **3a** in 57% yield. The yield of 1-(1'-fluoro-5'-pivaloxypentan-2'-yl)-cytosine **4** was 56% together with 22% of the less polar O<sup>2</sup>-isomer **4a**. The position of alkylation of the pyrimidine nucleus in each case was decided by comparison of NMR and UV measurements with literature values.<sup>10</sup> Although there was a marked difference in the R<sub>f</sub> values of the N- and O-isomers of the pyrimidine analogues, repeated flash chromatography failed to give a satisfactory separation. The pure isomers were isolated by preparative TLC. Removal of the pivaloyloxy group in **3** and **4** by refluxing in methanol with potassium carbonate (followed by acid hydrolysis of the 4-methoxy group in **3**), afforded compounds **7** and **8**.

The mixture resulting from the alkylation of adenine was separated by flash column chromatography. The main fraction (64%), which showed a single spot on TLC, was separated from an impure more polar fraction (16%). The main fraction was tentatively identified as the N-9 isomer **5** on the basis of <sup>13</sup>C NMR evidence. The C-2' signal appeared as a doublet at a higher field 54.9ppm, than the corresponding doublet at 61.5 ppm for the more polar fraction. **5a**. The pivaloyloxy protecting group was removed by refluxing with potassium carbonate in methanol. The N-9 site of alkylation in compound **9** was unambiguously confirmed by <sup>13</sup>C NMR<sup>11</sup>. Irradiation of the proton signal at 8.24 ppm showed decoupling with both the C-5 signal at 118.8 ppm<sup>12</sup> and the C-4 signal at 149.6 ppm. Irradiation of the signal at 8.15 showed decoupling with C-4 and with C-6 at

TABLE I <sup>1</sup> H NMR Spectral Data <sup>a</sup>							
Compd	Base Protons	H <sub>A</sub> -1'	H <sub>B</sub> -1'	H-2'	H <sub>2</sub> -3' H <sub>2</sub> -4'	H <sub>2</sub> -5'	Other
3 <sup>c</sup>	7.31 s	4.7, ddd J,47.1,10.3,3.4	4.63 ddd J,48.3,10.3,2.3	4.97 dm J,31.7	1.91 m 1.68 m	4.09 t J,6.3	4.00 (s,3H, OMe) 1.97 (d,3H, J,0.8 Me) 1.2 (s,9H,OMe <sub>3</sub> )
3a <sup>c</sup>	7.96 s	4.63 ddd J,50.0,10.0,5.0	4.58 ddd J,50.0,10.0,5.0	5.39 dm J,18.7	1.80 m	4.10 dt J,5.9	3.99 (s,3H,OMe) 1.94 (s,3H,Me) 1.19 (s,9H,Me <sub>3</sub> )
4 <sup>m</sup>	7.60 d 5.71d J,7.2 J,7.2	b	b 4.76 - 4.33 (3H,m)	b	1.70 m 1.47 m	3.98 t J,6.3	7.13 (bd, 2H,J,16.0,NH <sub>2</sub> ) 1.13 (s,(9H, CMe <sub>3</sub> )
4a <sup>m</sup>	7.84d 6.08d J,5.7 J,5.7	4.58, ddd J,47.6,10.3,3.4	4.54, ddd J,47.2,10.3,5.2	5.21 dm J,22.8	1.67 m	4.02 t J,5.6	6.87 (bs,2H,NH <sub>2</sub> ) 1.12 (s,9H,CMe <sub>3</sub> )
5 <sup>c</sup>	8.33 s 7.91 s	b	b 4.98 - 4.61 (3 H,m)	b	2.13 dm 1.63 dm	4.08 t J,5.7	6.10 (s,2H,NH <sub>2</sub> ) 1.18 (s,9H,CMe <sub>3</sub> )
5a <sup>c</sup>	8.13 s 8.02 s	5.11, ddd J,47.1,10.1,5.7	4.78, ddd J,47.2,10.1,2.7	4.98 dm J,27.0	2.28 dm 1.66 dm	4.08 t J,5.1	6.9 (bs,NH <sub>2</sub> ) 1.17 (s,9H,CMe <sub>3</sub> )
7 <sup>m</sup>	7.57 d J,0.9 11.28 bs	b	b 4.73 - 4.47 (4H,m) 3H after D <sub>2</sub> O	b	1.70 m 1.35 m	3.38 t J,6.3	1.79 (d,3H,J,0.9,Me) Obscured OH
8 <sup>m</sup>	7.57 d 5.70 d J,7.3 J,7.3	b	b 4.74 - 4.41 (4H,m) 3H after D <sub>2</sub> O	b	1.70 m 1.32 m	3.37 t J,6.3	7.04 (bd,2H,J,13.2,NH <sub>2</sub> ) Obscured OH
9 <sup>m</sup>	8.24 s 8.15 s	b	b 5.04 - 4.66 (3H,m)	b	1.95 m 1.29 dm	3.37 t H,6.3	7.26 (s,2H,NH <sub>2</sub> ) 4.45 (t,1H,J,5.1,OH)

<sup>a</sup>Signals are quoted as s (singlet), d (doublet), bs (broad singlet), bd (broad doublet), ddd double (double-doublet), t (triplet), dt (apparent double triplet), b Signal overlap. <sup>c</sup>In CDCl<sub>3</sub>, <sup>m</sup>In DMSO-d<sub>6</sub>.

TABLE II <sup>1</sup> H NMR SPECTRAL DATA <sup>a</sup>							
Compd	Base Protons	H <sub>A</sub> -S'	H <sub>B</sub> -S'	H-4'	H <sub>2</sub> -2'	H <sub>3</sub> -3'	Other
10 <sup>c</sup>	7.18 s J,1.1	4.47 ddd J,47.3,10.3,3.4	4.42 ddd J,47.3,10.3,4.8	5.08 dm J,31.0	1.88- 1.63m	3.84 m	3.98 (s,3H,OMe) 2.11 (s,3H,Ac) 1.95(d,3H,J1.08,Me)
10a <sup>c</sup>	7.98 d J,0.9	4.49 ddd J,47.4,10.2,3.2	4.44 ddd J,47.4,10.2,5.0	5.10 dm J,18.0	1.85 m	4.34 t J,6.2	3.98 (s,3H,OMe) 2.10 (s,3H,Ac) 2.05 (d,3H,J,0.93,Me)
11 <sup>m</sup>	8.15 8.14	4.49 ddd J,47.3,10.4,3.0	4.43 ddd J,47.3,10.4,5.2	5.01 dm J,21.3	1.84 m 1.53 m	4.16 t J,6.9	7.17 (s,2H,NH <sub>2</sub> ) 2.03 (s,3H,Ac)
13 <sup>m</sup>	7.82 d J,1.0	4.26 ddd J,47.6,9.3,4.2	4.21 ddd J,47.9,9.3,5.7	3.55 dm J,24.0	1.65 dm 1.33 m	3.74, t J,6.9	5.05 (bs,1H,OH) 3.83 (s,3H,OMe) 1.87 (d,3H,Me)
14 <sup>m</sup>	7.54 d J,1.2 11.21 bs,1H	4.27 ddd J,47.5,9.3,4.2	4.22 ddd J,47.9,9.3,5.6	b 3.63 m 3H	1.63 m 1.33 m	b 3.63 m 3H	5.04 (bs,1H,OH) 1.75 (d,3H,J,1.2)
15 <sup>m</sup>	7.57 d 5.63d J,7.1 J,7.1	4.27 ddd J,47.5,9.3,4.1	4.21 ddd 47.9,9.3,5.7	b 3.63 m	1.63 dm 1.32 m	b 3.63 m	6.97 (bd,2H,J,18.8) 5.00 (d,1H,J 5.4 OH)
15a <sup>m</sup>	7.84 d 6.07d J,5.8 J,5.8	4.29 ddd J,53.6,9.3,4.2	4.24 ddd J,53.6,9.3,5.7	3.65 m	1.8, m 1.4 m	4.15 t J,6.3	6.86 (bs,2H,NH <sub>2</sub> )
16 <sup>m</sup>	8.15 s 8.14 s	b 4.35 - 4.10	b 4.35 - 4.10	3.66 dm J,19.5	1.90 dm 1.30 m	b 4.35-4.10	7.20 (s,2H,NH <sub>2</sub> ) 5.03 (d,1H,OH)
17 <sup>m</sup>	7.70 s 10.62 s	4.27 ddd J,47.6,9.4,4.1	4.23 ddd J,47.9,9.4,5.6	3.65 dm J,19.0	1.60 dm 1.35 m	3.95 t J,6.1	6.50 (2H,NH <sub>2</sub> ) 5.0 (d,1H,OH)
(a) Signals are quoted as s (singlet), d (doublet), bs (broad singlet), bd (broad doublet) ddd (double double-doublet), t (triplet), dt (apparent double triplet), b Signal overlap <sup>1</sup> In CDCl <sub>3</sub> , <sup>3</sup> In DMSO-d <sub>6</sub>							

TABLE III <sup>13</sup> C NMR SPECTRAL DATA <sup>a</sup>									
Compd	C=O	Aromatic	C-5'	C-1'	C-2'	C-3'	C-4'	C-5'	Other
3 <sup>c</sup>	178.3	170.3 156.6 141.2, d J, 3.5	104.9	84.2 d J, 172	55.8 d J, 21.6	25.6 d J, 4.7	25.1	63.2	54.5 (OMe) 38.7 (CMe <sub>3</sub> ) 27.1 (Me) 12.2. (d, J, 1.7 Me)
3a <sup>c</sup>	178.4	169.7 163.0 156.8, d J, 1.2	111.4	83.5 d J, 172	73.8 d J, 19.8	26.6 d J, 5.3	24.4	63.8	53.9 (OMe), 38.6 (CMe <sub>3</sub> ) 27.1 (Me) 11.7 (d, J, 1.2, Me)
4 <sup>m</sup>	177.4	165.3 156.0 143.0	94.0	83.9 d J, 168	55.0 d J, 18.8	24.7 J, 4.7	24.6	63.3	38.8 (CMe <sub>3</sub> ) 26.9 (Me)
4a <sup>m</sup>	177.5	165.5 164.4 156.2	99.6	83.8 d J, 168.8	72.6 d J, 18.6	26.0 d J, 5.8	24.1	63.7	38.8 (CMe <sub>3</sub> ) 26.9 (Me)
5 <sup>c</sup>	178.3	155.6 152.8 149.8 139.3	119.5	83.6 d J, 173.8	54.9 d J, 19.2	26.5 J, 5.3	25.1	62.9	38.6 (CMe <sub>3</sub> ) 27.1 (Me)
5a <sup>c</sup>	178.3	154.7 153.4 150.0 141.5	121.0	82.5 d J, 173.8	61.5 d J, 18.7	25.2 d J, 5.3	25.1	62.8	38.6 (CMe) 27.0 (Me)
7 <sup>m</sup>		163.7 151.4 137.7	109.3	83.3 d J, 168	54.4 d J, 19.3	24.1 d J, 5.8	28.4	59.9	12.1 (Me)
8 <sup>m</sup>		165.2 155.5 143.0	93.7	84.0 d J, 167.0	55.3 d J, 19.0	24.7 J, 5.3	28.5	60.0	
9 <sup>m</sup>		156.0 152.4 149.6 139.9 d d bd bd <sup>3</sup> J <sup>1</sup> J <sup>3</sup> J <sup>1</sup> J 11.0 209.0 13.0 223	118.8 bd <sup>3</sup> J 8.0	83.67 d J, 169.0	54.8 d J, 18.0	25.5 d J, 5.8	28.6 d	59.9	
<sup>a</sup> Signals are quoted as d(doublet), bd (broad doublet) dm (double multiplet) <sup>1</sup> In CCl <sub>3</sub> <sup>m</sup> In DMSO-d <sub>6</sub>									

(continued)

TABLE III <sup>13</sup> C NMR SPECTRAL DATA <sup>a</sup>										
Compd	C=O	Aromatic	C-5'	C-1'	C-2'	C-3'	C-4'	C-5'	Other	
10 <sup>m</sup>	170.0	169.8 155.3 146.6	102.3	48.4	24.2	25.9 d J,6.4	71.4 d J,18.0	83.6 d J,167	53.9 (OMe) 20.7 (Ac) 11.5 (Me)	
10a <sup>m</sup>	170.1	167.0 163.2 157.2 d J,2.9	110.5	66.2	24.2	25.7 d J,6.4	71.5 d J,18.1	83.7 d J,168.5	55.9 (OMe) 20.1 (Ac) 11.5 (Me)	
11 <sup>m</sup>	170.0	155.9 152.3 149.5 140.8	118.7	42.9	25.2	26.0 J,6.4	71.2 d J,17.5	83.6 d J,168.5	20.7 (Ac)	
13 <sup>m</sup>		168.9 163.3 157.2	110.3	66.7	24.6	28.7 d J,6.4	68.3 d J,18.7	86.9 d J,168.0	53.7 (OMe) 11.5 Me	
13a <sup>m</sup>		170.0 156.0 146.4	103.3	49.3	25.0	29.1 d J,6.4	68.6 d J,18.7	86.9 d J,167.4	54.4 (OMe) 11.8 (Me)	
14 <sup>m</sup>		164.3 150.9 141.5	108.41	47.1	24.7	28.9 d J,5.8	68.2 d J,18.7	86.7 d J,168.0	12.0 (Me)	
15 <sup>m</sup>		165.9 155.9 146.0	93.1	48.5	25.0	29.0 d J,5.8	68.3 d J,18.4	86.7 d J,166.8		
15a <sup>m</sup>		165.4 164.9 156.2	99.3	65.7	24.7	28.8 d J,6.4	68.3 d J,18.3	86.6 d J,167		
16 <sup>m</sup>		156.0 152.4 149.6 140.9 d d dm bd <sup>3</sup> J <sup>1</sup> J <sup>3</sup> J <sup>1</sup> J	118.8 bd <sup>3</sup> J	42.9	25.7	29.1 d J,7.0	68.1 d J,18.7	86.6 d J,168		
17 <sup>m</sup>		156.7 153.4 151.1 137.4	116.5	42.6	25.6	29.1 d J,6.3	68.0 d J,18.9	86.5 d J,168.7		
<sup>a</sup> Signals are quoted as d (doublet), bd (broad doublet) dm (double multiplet) <sup>c</sup> In CDCl <sub>3</sub> <sup>m</sup> In DMSO-d <sub>6</sub>										

156.0 ppm. A 2-D NMR  $^1\text{H}$ - $^{13}\text{C}$  correlation experiment connected the signals 8.24 (8-H) with 139.9 (C-8) and 8.15 (2-H) with 152.4 (C-2). The C-8 signal was upfield from the C-2 signal which is characteristic of N-9 alkylated adenines.<sup>11</sup> Irradiation at 4.88 ppm showed decoupling with C-4 and C-8. For compound **16** the separation between signals 8-H and 2-H was too small for selective irradiation. Decoupling at 4.16 ppm showed correlation with C-4 at 149.6 ppm and the signal at 140.9 ppm. Compound **16** was assigned the N-9 structure on the basis of literature values<sup>13</sup> and the similarities of the UV and  $^{13}\text{C}$  NMR data with compound **9**.

Condensation of the iodide with nucleic acid bases by stirring with potassium carbonate in DMF at ambient temperature for 24h, gave compounds **10**, **11**, **13** and **16**. Cesium carbonate replaced potassium carbonate in the reaction with cytosine which gave the N- and O- isomers of **15** directly. Hydrolysis with 10% HCl of compounds **10** and **12** gave **14** and **17** respectively.

### Evaluation of Antiviral Activity

The antiviral activity and cytotoxicity of the compounds are shown in Table IV. All the compounds were inactive against all viruses, except for compound **15** which showed a modest activity against cytomegalovirus ( $\text{MIC}_{50}$ : 12-15  $\mu\text{g/ml}$ ). This anti-CMV activity must be considered as specific since (1) compound **15** was not inhibitory to the host cell growth unless its concentration was raised to 200  $\mu\text{g/ml}$  and (11) compound **15** did not show activity against any of the other viruses tested.

### Experimental

M.p.s. were determined on a Gallenkamp capillary apparatus and are uncorrected; UV spectra were obtained on a Unicam SP800A spectrometer. NMR spectra Tables I-III were recorded in ppm on a Bruker MSL300 machine at 300.13 MHz for  $^1\text{H}$  and 75.468 MHz for  $^{13}\text{C}$ .  $\text{SiMe}_4$  was the internal standard, J values are given in Hz. TLC was carried out on merck silica gel 60F<sub>254</sub>-coated aluminium sheets and spots were visualised by UV illumination. Column chromatography was carried out on Merck silica gel 60 (230-400 mesh) or 60 (70-230 mesh).



TABLE IV

Virus (strain)	Cell	MIC <sub>50</sub> <sup>ab</sup> , (μg/mL)								15	Ribavirin	Ganciclovir	Acyclovir	BVDU
		9	7	8	17	16	14	15						
HSV-1 (KOS)	ESM	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	0.002	0.07	0.001	
HSV-2 (G)	ESM	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	0.007	0.07	200	
TK <sup>-</sup> HSV-1 (B2006)	ESM	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	100	20	70	100	
VZV (Oka)	HEL	> 50	> 50	> 50	> 50	> 50	> 50	> 50	> 50			0.13	0.0001	
VZV (YS)	HEL	> 50	> 50	> 50	> 50	> 50	> 50	> 50	> 50			0.27	0.0002	
TK <sup>-</sup> VZV (07-01)	HEL	> 50	> 50	> 50	> 50	> 50	> 50	> 50	> 50			9	> 50	
TK <sup>-</sup> VZV (YS-R)	HEL	> 50	> 50	> 50	> 50	> 50	> 50	> 50	> 50			9	> 50	
CMV (AD-169)	HEL	> 50	> 50	> 50	> 50	> 50	> 50	> 50	15		0.7			
CMV (Davis)	HEL	> 50	> 50	> 50	> 50	> 50	> 50	> 50	12		0.5			
Vaccinia	ESM	> 400	> 200	> 400	> 400	> 400	> 400	> 400	> 400	40	> 100	> 400	0.2	
Polio-1	HeLa	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	70			> 400	
Coxsackie B4	HeLa	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	70			> 400	
Reo-1	Vero	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	150			> 400	
Parainfluenza-3	Vero	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	70			> 400	
Sindbis	Vero	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	70			> 400	
Semliki forest	Vero	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	200			> 400	
Junin	Vero	> 50	> 200	> 50	> 200	> 50	> 200	> 200	> 200	8				
Tacaribe	Vero	> 50	> 200	> 50	> 200	> 50	> 200	> 200	> 200	5				
Vesicular stomatitis	ESM	> 400	> 400	> 400	> 400	> 400	> 400	> 400	> 400	10	> 100	> 400	> 400	
HIV-1 (IIIB/LAI)	CEM	> 100	> 100	> 100	> 100	> 100	> 100	> 100	> 100					
HIV-2(ROD)	CEM	> 100	> 100	> 100	> 100	> 100	> 100	> 100	> 100					
Cell growth	HEL				150	100	150	200			> 50	> 200	> 200	

<sup>a</sup>Minimal inhibitory concentration required to reduce virus-induced cytopathicity by 50%. Abbreviations for viruses and cells; HSV-1, herpes simplex virus type 1; HSV-2, herpes simplex virus type 2; TK<sup>-</sup>, thymidine kinase-deficient; VZV, varicella-zoster virus; CMV, cytomegalovirus; HIV-1, human immunodeficiency virus type 1; HIV-2, human immunodeficiency virus type 2; ESM, (human) embryonic-skin muscle (cells); HEL, human embryonic lung (fibroblasts); HeLa, human cervix carcinoma (epithelial) cells; Vero, African green monkey kidney cells; CEM, human T-4 lymphocyte cells.

<sup>b</sup>None of the compounds proved cytotoxic to the cells, as monitored by microscopically visible alteration of normal cell morphology, at the highest concentrations tested.

1-(1'-Fluoro-5'-pivaloyloxypentan-2'-yl)-O<sup>4</sup>-methylthymine **3** The tosyl compound **2** (550 mg, 1.5 mmol) was added to a mixture of 4-methoxy-5-methyl-2-pyrimidinone (240 mg, 1.65 mmol), potassium carbonate (700 mg, 5.0 mmol) and 18-crown-6 (370 mg, 1.5 mmol) in DMF (5 cm<sup>3</sup>) and stirred at 65°C for 72h. The solvent was evaporated. The residue was triturated with 2 x 50 cm<sup>3</sup> of ethyl acetate, filtered through a pad of silica gel and evaporated. TLC of the crude oil using dichloromethane - ethyl acetate (10:1) gave the O<sup>2</sup>-isomer **3a**, R<sub>f</sub> 0.68, as an oil (283 mg, 57%); UV (MeOH) λ<sub>max</sub> 268 nm. The title compound **3**, R<sub>f</sub> 0.34, (149 mg, 30%) was crystallised from hexane-diethyl ether (1:1) to give needles, m.p. 94°C; UV(MeOH) λ<sub>max</sub> 284 nm. Anal. (C<sub>16</sub>H<sub>25</sub>FO<sub>4</sub>N<sub>2</sub>) calculated: C, 58.52; H, 7.67; N, 8.53. Found: C, 58.52; H, 7.46; N, 8.38.

1-(1'-Fluoro-5'-pivaloyloxypentan-2'-yl)-cytosine **4**. A mixture of the tosyl ester **2** (360 mg, 1 mmol) cytosine (116 mg, 1.05 mmol) and cesium carbonate (655 mg, 2 mmol) in dry DMF (5 cm<sup>3</sup>) was stirred at 75°C for 17 h. The solvent was evaporated. The residue was triturated with ethyl acetate-methanol (5:1, 2 x 50 cm<sup>3</sup>), filtered through a pad of silica gel and evaporated. TLC of the crude oil with ethyl acetate gave the O<sup>2</sup>-isomer **4a**, R<sub>f</sub> 0.85, as an oil (66 mg, 22%). UV (MeOH) λ<sub>max</sub> 272 nm. UV (0.01 M HCl) 262 nm. The title compound **4** R<sub>f</sub> 0.35, (170 mg, 56%), after trituration with pentane-acetone 1:1, gave a white powder. UV (MeOH) λ<sub>max</sub> 275 nm. UV (0.01 HCl) λ<sub>max</sub> 285 nm. Anal. (C<sub>14</sub>H<sub>22</sub>FN<sub>3</sub>O<sub>3</sub>): calculated C, 56.17; H, 7.41; N, 14.04. Found: C, 55.98; H, 7.17; N, 13.89.

9-(1'-Fluoro-5'-pivaloyloxypentan-2'-yl)-9H-adenine **5**. The tosyl ester **2** was condensed with adenine in the same way as described for compound **3**. The crude product was flash chromatographed twice. Elution with dichloromethane-methanol (100:5) gave the title compound **5** (208 mg, 64%) which was recrystallised from dichloromethane-hexane, (1:1). m.p. 125-126°C. UV (MeOH) λ<sub>max</sub> 262 nm. Anal. (C<sub>15</sub>H<sub>22</sub> F N<sub>5</sub> O<sub>2</sub>): Calculated C, 55.72; H, 6.86; N, 21.66. Found: C, 55.58; H, 6.78; N, 21.80.

Elution of the column with dichloromethane-methanol (100:7) gave the impure N-3 isomer **5a** as a yellow gum. λ<sub>max</sub> (MeOH) 277 nm.

1-(1-Fluoro-5'-hydroxypentan-2'-yl)-thymine 7. A mixture of the ester **3** (75 mg, 0.23 mmol) and anhydrous potassium carbonate (200 mg, 1.5 mmol) in methanol (5 cm<sup>3</sup>) was refluxed with stirring for 3 h. The mixture was acidified to pH2 with 10% HCl, stirred at ambient temperature for 1 hr and evaporated. The residue was co-evaporated with ethanol (3 x 5 cm<sup>3</sup>) and triturated with ethanol (5 cm<sup>3</sup>). The ethanol soluble material was purified by TLC using ethyl acetate - methanol (10:1). Elution with ethanol gave a clear oil (40 mg, 76%) which on trituration with diethyl ether gave a white solid. UV (MeOH)  $\lambda_{\text{max}}$  270 nm. Anal. (C<sub>10</sub>H<sub>15</sub>FN<sub>2</sub>O<sub>3</sub> 0.25 H<sub>2</sub>O): calculated: C, 51.17; H, 6.66; N, 11.93; found: C, 51.44; H, 6.43; N, 11.65.

1-(1'-Fluoro-5'-hydroxypentan-2'-yl)-cytosine 8 A mixture of compound **4** (184 mg, 0.62 mmol) and anhydrous potassium carbonate (500 mg, 3.6 mmol) in methanol (5 cm<sup>3</sup>) was refluxed with stirring for 3h. The mixture was acidified to pH7 with 10% HCl and the solvent removed *in vacuo*. The residue was co-evaporated with ethanol (3 x 5 cm<sup>3</sup>) and triturated with ethanol. The ethanol soluble material was purified by TLC with ethanol, to give the title compound **8** (55 mg, 41%). Trituration with ethanol-diethyl ether gave a white powder. UV (MeOH)  $\lambda_{\text{max}}$  274 nm. Anal. (C<sub>9</sub>H<sub>14</sub>FN<sub>3</sub>O<sub>2</sub> 0.5 H<sub>2</sub>O). Calculated: C, 48.21; H, 6.74; N, 18.74. Found C, 48.29; H, 6.43; N, 18.60.

9-(1'-Fluoro-5'-hydroxypentan-2'-yl)-adenine 9. Compound **5** (110 mg, 0.34 mmol) was hydrolysed as described for compound **8**. The ethanol soluble material was flash chromatographed. Elution with dichloromethane-methanol (100:16) gave a clear gum which was triturated with diethyl ether to yield the title compound **9** (58 mg, 71%) as a white powder. UV (MeOH)  $\lambda_{\text{max}}$  263 nm. Anal. (C<sub>10</sub>H<sub>14</sub>N<sub>5</sub>OF 0.2H<sub>2</sub>O). Calculated: C, 49.46; H, 5.98; N, 28.84. Found: C, 49.10; H, 5.97; N, 28.53.

1-(4'-acetoxy-5'-fluoropentyl)-O<sup>4</sup>-methylthymine 10. The iodide **1** (276 mg, 1 mmol) was added to a mixture of 4-methoxy-5-methyl-2-pyrimidinone (140 mg, 1 mmol) and anhydrous potassium carbonate (276 mg, 2.0 mmol) in DMF (5 cm<sup>3</sup>) and stirred at ambient temperature for 24h. The solvent was evaporated, the residue triturated with ethanol (2 x 50 cm<sup>3</sup>) and the ethanol solutions filtered through a pad of silica gel (10 g).

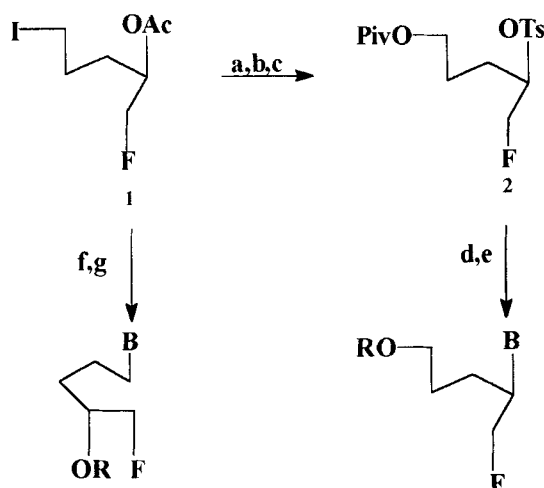
Evaporation of the combined filtrate left an oil which was separated by TLC with dichloromethane-ethyl acetate (10:1) to give the O<sup>2</sup>-isomer of **10a** R<sub>f</sub> 0.63 (28 mg, 10%) UV (MeOH) λ<sub>max</sub> 266 nm and the title compound **10** R<sub>f</sub> 0.13 (184 mg, 64%), as a colorless gum, UV (MeOH) λ<sub>max</sub> 284 nm. Anal. (C<sub>13</sub>H<sub>19</sub>FN<sub>2</sub>O<sub>4</sub>·0.5H<sub>2</sub>O). Calculated: C, 52.88; H, 6.83; N, 9.49. Found: C, 52.86; H, 6.57; N, 9.14.

Partial hydrolysis of the acetate group occurred during work up. 1-(5'-fluoro-4'-hydroxypentyl)-O<sup>4</sup>-methylthymine **13** was isolated by TLC with dichloromethane-methanol (2:1) and recrystallised from ethanol, m.p. 111–113°C. UV (MeOH) λ<sub>max</sub> 283 nm. Anal. (C<sub>11</sub>H<sub>17</sub>FN<sub>2</sub>O<sub>3</sub>·0.5H<sub>2</sub>O). Calculated: C, 52.16; H, 7.16; N, 11.06. Found: C, 52.15; H, 6.90; N, 10.94.

1-(5'-Fluoro-4'-hydroxypentyl)-thymine 14. The acetate **10** (143 mg, 0.5 mmol) was refluxed for 1 h in ethanol: 10% HCl (1:1). The cooled solution was adjusted to pH7 with sodium carbonate and the solvent evaporated *in vacuo*. The residue was co-evaporated with ethanol (3 x 5 cm<sup>3</sup> and then triturated with ethanol (10 cm<sup>3</sup>). TLC of the ethanol soluble material with ethyl acetate-methanol (10:1) gave the title compound **14** as a white powder. (72 mg, 63%). UV (MeOH) λ<sub>max</sub> 272 nm. Anal. (C<sub>10</sub>H<sub>15</sub>FN<sub>2</sub>O<sub>3</sub>·0.5H<sub>2</sub>O). Calculated: C, 50.20; H, 6.74; N, 11.71. Found: C, 50.35; H, 6.51; N, 11.34.

1-(5'-Fluoro-4'-hydroxypentyl)-cytosine 15. Cytosine (1 mmol) was alkylated with the iodide (1 mmol) using the same procedure as described for compound **10** except that anhydrous cesium carbonate (652 mg, 2 mmol) replaced potassium carbonate. TLC of the crude product with ethanol gave the O<sup>2</sup>-isomer **15a**. R<sub>f</sub> 0.85 (37 mg, 17%). UV (MeOH) λ<sub>max</sub> 272 nm. UV (0.01 HCl) λ<sub>max</sub> 261 nm. The title compound **15** R<sub>f</sub> 0.54 (157 mg, 73%) after trituration with ethanol-diethyl ether was a white powder. UV (MeOH) λ<sub>max</sub> 273 nm. UV (0.01 HCl) λ<sub>max</sub> 288 nm. Anal. (C<sub>9</sub>H<sub>14</sub>FN<sub>3</sub>O<sub>2</sub>·0.5H<sub>2</sub>O). Calculated: C, 48.21; H, 6.74; N, 18.74. Found: C, 47.98; H, 6.33; N, 18.48.

9-(5'-Fluor-4-hydroxypentan-1'-yl)-9H-adenine 16 and 9-(4'-acetoxy-5'-fluoropentyl)-9H-adenine 11. Adenine (135 mg, 1 mmol) was alkylated with the iodide as described for compound **10**. The crude product was flash chromatographed. Elution with dichloromethane-methanol (10:1) gave the acetate (35 mg, 12%) which crystallised from



10. R=Ac, B=O<sup>4</sup>-methylthymine-1-yl

11. R=Ac, B=adenine-9-yl

12. R=Ac, B=2-amino-6-chloropurine-9-yl

13. R=H, B=O<sup>4</sup>-methylthymine-1-yl

14. R=H, B=thymine-1-yl

15. R=H, B=cytosine-1-yl

16. R=H, B=adenine-9-yl

17. R=H, B=guanine-9-yl

3. R=Piv, B=O<sup>4</sup>-methylthymine-1-yl

4. R=Piv, B=cytosine-1-yl

5. R=Piv, B=adenine-9-yl

6. R=Piv, B=2-amino-6-chloropurine-9-yl

7. R=H, B=thymine-1-yl

8. R=H, B=cytosine-1-yl

9. R=H, B=adenine-9-yl

a. Me<sub>3</sub>CCO<sub>2</sub>SnBu<sub>3</sub>, CsF, DMF 40°C; b. NH<sub>3</sub>-MeOH, ambient temp.; c. (Bu<sub>3</sub>Sn)<sub>2</sub>O, toluene; then p-TsCl, DMAP, Et<sub>3</sub>N, 65°C. d. 5-methyl-4-methoxy-2 pyrimidinone or adenine, K<sub>2</sub>CO<sub>3</sub>, 18-C-6, DMF, 65°, or cytosine, Cs<sub>2</sub>CO<sub>3</sub> DMF, 75°C; e. K<sub>2</sub>CO<sub>3</sub>, MeOH, 65°C. f. 5-methyl-4-methoxy-2-pyrimidinone or adenine K<sub>2</sub>CO<sub>3</sub>, DMF ambient temp., or cytosine, Cs<sub>2</sub>CO<sub>3</sub>, DMF, ambient temp.; g. 2M HCl.

ethanol as needles m.p. 143 -145°C. UV (MeOH)  $\lambda_{\max}$  265 nm. Anal. ( $C_{12}H_{16}FN_3O_2 \cdot 0.25H_2O$ ). Calculated: C, 50.93; H, 5.82; N, 24.56. Found: C, 50.64; H, 5.66; N, 24.59. Elution with dichloromethane methanol (7:1) gave the title compound **16** (172 mg, 69%) as a white powder. UV (MeOH)  $\lambda_{\max}$  261. Anal. ( $C_{10}H_{14}FN_3O \cdot 0.5H_2O$ ). Calculated: C, 48.38; H, 6.09; N, 28.21. Found: C, 48.79; H, 5.97; N, 27.91.

9-(5'-fluoro-4'-hydroxypentyl)-9H-guanine **17** Compound **12** (126 mg, 0.34 mmol) was refluxed for 2 hr with 10% HCl. (5cm<sup>3</sup>). The mixture was cooled, adjusted to pH7 with sodium carbonate solution and filtered. Recrystallisation of the solid from water gave the title compound m.p. 185-187°C) (93 mg, 83%). UV (H<sub>2</sub>O)  $\lambda_{\max}$  253 nm. Anal. ( $C_{10}H_{14}FN_3O_2 \cdot 1.5 H_2O$ ). Calculated: C, 42.55; H, 6.08; N, 24.80. Found: C, 42.38; H, 5.79; N, 24.53.

### Viruses and Antiviral Assays

The source of the viruses and methodology of the antiviral assays are described in previous publications for herpes simplex virus<sup>14</sup>, and most other viruses<sup>15</sup>, varicella zoster and cytomegalovirus<sup>16</sup>, human immunodeficiency virus<sup>17</sup> and the myxoviruses<sup>18</sup>

### Acknowledgements

We are grateful to Dr. John O'Brien for NMR spectra. The work was supported in part by the AIDS Basic Research Programme of the European Community, and grants from the Belgian National Fonds voor Wetenschappelijk Onderzoek and the Belgian Fonds voor Geneeskundig Wetenschappelijk Onderzoek. We thank Dr. J. Balzarini, Dr. R. Snoeck, Dr. G. Andrei and Dr. S. Ikeda as well as Mrs. A. Van Lierde, Mrs. F. De Meyer, Mrs. A. Camps and Mrs. A. Absillis for help with the evaluation of the antiviral activity.

### References

1. Schaeffer, H.J.; Beauchamp, L.; de Miranda, P.; Elion, G.B.; Bauer, D.J. Collins, P. *Nature*, **1978**, 272, 583.

2. For reviews, see: Chu, C.K.; Cutler, S.J. *J. Heterocycl. Chem.*, **1986**, *23*, 289; Remy R.J.; Secrist, J.A. *Nucleosides, Nucleotides*, **1985**, *4*, 411.
3. El-Kattan Y.; Gosselin, G.; Imbach, J.L. *J.Chem.Soc., Perkin Trans. 1*, **1994**, 1289 and references cited therein.
4. Bricand, H.; Herdewijn, P.; De Clercq, E. *Biochem. Pharmacol.*, **1983**, 3583
5. (a) Mann, J. *Chem.Soc.Rev.*, **1987**, *16*, 381.  
(b) Abeles, R.H.; and Alston, T.A. *J.Biol.Chem.*, **1990**, *265*, 16705.
6. Pankiewicz, K.W.; Krzeminski, J.; Ciszewski, L.A.; Rem, W.; Watanabe, K.A. *J.Org.Chem.*, **1992**, *57*, 553 and references cited therein.
7. Lewis, M.; McMurry, T.B.H.; De Clercq, E. *J. Chem. Soc. Perkin Trans. 1*, **1993**, 2107-2110.
8. Medich, J.R.; Kunnen, K.B. Johnson, C.R.; *Tetrahedron Lett.*, **1987**, *28*, 4131-4134.
9. Bronson, J.J.; Ghazzouli, I.; Hitchcock, M.J.M.; Webb II, R.R.; Martin, J.C. *J. Med. Chem.*, **1989**, *32*, 1457-1463.
10. Vandendriessche, F.; Snoeck, R.; Janssen, G.; Hoogmartens, J.; Van Aerschot, A.; De Clercq, E.; Herdewijn, P. *J. Med. Chem.*, **1992**, *35*, 1458-1465.
11. Platzer, N.; Galons, H.; Bensaid, Y.; Miocque, M.; Bram, G. *Tetrahedron*, **1987**, *43*, 2101-2108.
12. Chenon, M.T.; Pugmire, R.J.; Grant, D.M.; Panzica, R.P.; Townsend, L.B. *J. Am. Chem. Soc.*, **1975**, *97*, 4627-4636.
13. Townsend, L.B.; Robins, R.K.; Loeppky, R.N.; Leonard, N.J. *J. Am. Chem.Soc.*, **1964**, *86*, 5320-5325.

14. De Clercq, E.; Descamps, J.; Verhelst, G.; Walker, R.T.; Jones, A.S.; Torrence, P.F.; Shugar, D. *J.Infect.Dis.*, **1980**, *141*, 563.
15. De Clercq, E. *Antimicrob.Agents Chemother.*, **1985**, *28*, 84.
16. De Clercq, E.; Holy, A.; Rosenberg, I.; Sakum, T.; Balzarini J.; Maudgal, P.C. *Nature*, **1986**, *323*, 464.
17. Pauwels, R.; De Clercq, E.; Desmyter, J.; Balzarini, J.; Goubau, P.; Herdewijn, P.; Vanderhaeghe H.; Vandeputte, M. *J.Virol.Methods*, **1987**, *16*, 171.
18. Hosoya, M.; Balzarini, J.; Shigeta, S.; De Clercq, E. *Antimicrob. Agents Chemother.*, **1991**, *35*, 2515.

Received March 17, 1995

Accepted August 9, 1995